

# Package: dsm (via r-universe)

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**License** GPL (>= 2)

**Title** Density Surface Modelling of Distance Sampling Data

**LazyLoad** yes

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**Description** Density surface modelling of line transect data. A  
Generalized Additive Model-based approach is used to calculate  
spatially-explicit estimates of animal abundance from distance  
sampling (also presence/absence and strip transect) data.  
Several utility functions are provided for model checking,  
plotting and variance estimation.

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**Language** en-GB

**Encoding** UTF-8

**URL** <https://github.com/DistanceDevelopment/dsm>

**BugReports** <https://github.com/DistanceDevelopment/dsm/issues>

**Depends** R (>= 3.5.0), mgcv (>= 1.8-23), mrds (>= 2.1.16), numDeriv

**Imports** nlme, ggplot2, plyr, statmod

**Suggests** Distance, sp, tweedie, testthat

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**Repository** <https://distancedevelopment.r-universe.dev>

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`dsm-package`*Density surface modelling*

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## Description

dsm implements spatial models for distance sampling data. Models for detectability can be fitted using packages `mrds` or `Distance`. dsm fits generalized additive models to spatially-referenced data. See Miller et al (2013) for an introduction.

## Details

Further information on distance sampling methods and example code is available at <http://distancesampling.org/R/>.

For help with distance sampling and this package, there is a Google Group <https://groups.google.com/forum/#!forum/distance-sampling>.

A example analyses are available at <http://examples.distancesampling.org>.

## References

Hedley, S. and S. T. Buckland. 2004. Spatial models for line transect sampling. *JABES* 9:181-199.

Miller, D. L., Burt, M. L., Rexstad, E. A., Thomas, L. (2013), Spatial models for distance sampling data: recent developments and future directions. *Methods in Ecology and Evolution*, 4: 1001-1010. doi: 10.1111/2041-210X.12105 (Open Access)

Wood, S.N. 2006. *Generalized Additive Models: An Introduction with R*. CRC/Chapman & Hall.

---

`block.info.per.su`*Find the block information*

---

## Description

Takes the transect data and works out how many blocks of a given size (in segment terms) fit into each.

## Usage

```
block.info.per.su(block.size, data, name.su)
```

## Arguments

<code>block.size</code>	number of segments per block
<code>data</code>	data used to build the model
<code>name.su</code>	names of the sampling units (i.e., transects)

**Value**

a `data.frame` with the following columns

- `name` the sample unit name (e.g. transect label)
- `num.seg` number of segments in that transect
- `num.block` number of blocks available
- `start.block` block number for first block
- `end.block` block number for last block
- `num.req` number of blocks needed for the unit

---

`check.cols`

*Check column names exist*

---

**Description**

Internal function to check that supplied `data.frames` have the correct columns and checks that sample labels are all unique.

**Usage**

```
check.cols(ddf.obj, segment.data, observation.data, segment.area)
```

**Arguments**

`ddf.obj` a `ddf` object from [mrds](#)  
`segment.data` segment data as defined in [dsm](#)  
`observation.data` observation data as defined in [dsm](#)  
`segment.area` area of segments

**Value**

nothing, but throws an error if something went wrong

**Author(s)**

David Lawrence Miller

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dsm	<i>Fit a density surface model to segment-specific estimates of abundance or density.</i>
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## Description

Fits a density surface model (DSM) to detection adjusted counts from a spatially-referenced distance sampling analysis. `dsm` takes observations of animals, allocates them to segments of line (or strip transects) and optionally adjusts the counts based on detectability using a supplied detection function model. A generalized additive model, generalized mixed model or generalized linear model is then used to model these adjusted counts based on a formula involving environmental covariates.

## Usage

```
dsm(
  formula,
  ddf.obj,
  segment.data,
  observation.data,
  engine = "gam",
  convert.units = 1,
  family = quasipoisson(link = "log"),
  group = FALSE,
  control = list(keepData = TRUE),
  availability = 1,
  segment.area = NULL,
  weights = NULL,
  method = "REML",
  ...
)
```

## Arguments

<code>formula</code>	formula for the surface. This should be a valid formula. See "Details", below, for how to define the response.
<code>ddf.obj</code>	result from call to <code>ddf</code> or <code>ds</code> . If multiple detection functions are required a <code>list</code> can be provided. For strip/circle transects where it is assumed all objects are observed, see <code>dummy_ddf</code> . Mark-recapture distance sampling ( <code>mrds</code> ) models of type <code>io</code> (independent observers) and <code>trial</code> are allowed.
<code>segment.data</code>	segment data, see <a href="#">dsm-data</a> .
<code>observation.data</code>	observation data, see <a href="#">dsm-data</a> .
<code>engine</code>	which fitting engine should be used for the DSM (" <code>glm</code> "/" <code>gam</code> "/" <code>gamm</code> "/" <code>bam</code> ").
<code>convert.units</code>	conversion factor to multiply the area of the segments by. See 'Units' below.

family	response distribution (popular choices include <a href="#">quasipoisson</a> , <a href="#">Tweedie/tw</a> and <a href="#">negbin/nb</a> ). Defaults <a href="#">quasipoisson</a> .
group	if TRUE the abundance of <i>groups</i> will be calculated rather than the abundance of <i>individuals</i> . Setting this option to TRUE is equivalent to setting the size of each group to be 1.
control	the usual control argument for a <a href="#">gam</a> ; keepData must be TRUE for variance estimation to work (though this option cannot be set for GLMs or GAMMs).
availability	an estimate of availability bias. For count models used to multiply the effective strip width (must be a vector of length 1 or length the number of rows in <code>segment.data</code> ); for estimated abundance/estimated density models used to scale the response (must be a vector of length 1 or length the number of rows in <code>observation.data</code> ). Uncertainty in the availability is not handled at present.
segment.area	if NULL (default) segment areas will be calculated by multiplying the Effort column in <code>segment.data</code> by the (right minus left) truncation distance for the <code>ddf.obj</code> or by <code>strip.width</code> . Alternatively a vector of segment areas can be provided (which must be the same length as the number of rows in <code>segment.data</code> ) or a character string giving the name of a column in <code>segment.data</code> which contains the areas. If <code>segment.area</code> is specified it takes precedent.
weights	weights for each observation used in model fitting. The default, <code>weights=NULL</code> , weights each observation by its area (see Details). Setting a scalar value (e.g., <code>weights=1</code> ) all observations are equally weighted.
method	The smoothing parameter estimation method. Default is "REML", using Restricted Maximum Likelihood. See <a href="#">gam</a> for other options. Ignored for <code>engine="glm"</code> .
...	anything else to be passed straight to <a href="#">glm</a> , <a href="#">gam</a> , <a href="#">gamm</a> or <a href="#">bam</a> .

## Details

The response (LHS of formula) can be one of the following (with restrictions outlined below):

- `count` count in each segment
- `abundance.est` estimated abundance per segment, estimation is via a Horvitz-Thompson estimator
- `density.est` density per segment

The offset used in the model is dependent on the response:

- `count` area of segment multiplied by average probability of detection in the segment
- `abundance.est` area of the segment
- `density` zero

The count response can only be used when detection function covariates only vary between segments/points (not within). For example, weather conditions (like visibility or sea state) or foliage cover are usually acceptable as they do not change within the segment, but animal sex or behaviour will not work. The `abundance.est` response can be used with any covariates in the detection function.

In the density case, observations can be weighted by segment areas via the `weights=` argument. By default (`weights=NULL`), when density is estimated the weights are set to the segment areas

(using `segment.area` or by calculated from detection function object metadata and `Effort` data). Alternatively `weights=1` will set the weights to all be equal. A third alternative is to pass in a vector of length equal to the number of segments, containing appropriate weights.

A example analyses are available at <http://examples.distancesampling.org>.

### Value

a `glm`, `gam`, `gamm` or `bam` object, with an additional element, `$ddf` which holds the detection function object.

### Units

It is often the case that distances are collected in metres and segment lengths are recorded in kilometres. `dsm` allows you to provide a conversion factor (`convert.units`) to multiply the areas by. For example: if distances are in metres and segment lengths are in kilometres setting `convert.units=1000` will lead to the analysis being in metres. Setting `convert.units=1/1000` will lead to the analysis being in kilometres. The conversion factor will be applied to `segment.area` if that is specified.

### Large models

For large models, `engine="bam"` with `method="fREML"` may be useful. Models specified for `bam` should be as `gam`. Read `bam` before using this option; this option is considered EXPERIMENTAL at the moment. In particular note that the default basis choice (thin plate regression splines) will be slow and that in general fitting is less stable than when using `gam`. For negative binomial response, `theta` must be specified when using `bam`.

### Author(s)

David L. Miller

### References

- Hedley, S. and S. T. Buckland. 2004. Spatial models for line transect sampling. *JABES* 9:181-199.
- Miller, D. L., Burt, M. L., Rexstad, E. A., Thomas, L. (2013), Spatial models for distance sampling data: recent developments and future directions. *Methods in Ecology and Evolution*, 4: 1001-1010. doi: 10.1111/2041-210X.12105 (Open Access)
- Wood, S.N. 2006. *Generalized Additive Models: An Introduction with R*. CRC/Chapman & Hall.

### Examples

```
## Not run:
library(Distance)
library(dsm)

# load the Gulf of Mexico dolphin data (see ?mexdolphins)
data(mexdolphins)

# fit a detection function and look at the summary
hr.model <- ds(distdata, truncation=6000,
```

```

      key = "hr", adjustment = NULL)
summary(hr.model)

# fit a simple smooth of x and y to counts
mod1 <- dsm(count~s(x,y), hr.model, segdata, obsdata)
summary(mod1)

# predict over a grid
mod1.pred <- predict(mod1, preddata, preddata$area)

# calculate the predicted abundance over the grid
sum(mod1.pred)

# plot the smooth
plot(mod1)

## End(Not run)

```

---

 dsm-data

*Data format for DSM*


---

## Description

Two data.frames must be provided to `dsm`. They are referred to as `observation.data` and `segment.data`.

## Details

The `segment.data` table has the sample identifiers which define the segments, the corresponding effort (line length) expended and the environmental covariates that will be used to model abundance/density. `observation.data` provides a link table between the observations used in the detection function and the samples (segments), so that we can aggregate the observations to the segments (i.e., `observation.data` is a "look-up table" between the observations and the segments).

`observation.data` - the observation data.frame must have (at least) the following columns:

- `object` unique object identifier
- `Sample.Label` the identifier for the segment where observation occurred
- `size` the size of each observed group (e.g., 1 if all animals occurred individually)
- `distance` distance to observation

One can often also use `observation.data` to fit a detection function (so additional columns for detection function covariates are allowed in this table).

`segment.data`: the segment data.frame must have (at least) the following columns:

- `Effort` the effort (in terms of length of the segment)
- `Sample.Label` identifier for the segment (unique!)
- `???` environmental covariates, for example location (projected latitude and longitude), and other relevant covariates (sea surface temperature, foliage type, altitude, bathymetry etc).



### Multiple detection functions

If multiple detection functions are to be used, then a column named `ddfobj` must be included in `observation.data` and `segment.data`. This lets the model know which detection function each observation is from. These are numeric and ordered as the `ddf.obj` argument to `dsm`, e.g., `ddf.obj=list(ship_ddf, aerial_ddf)` means ship detections have `ddfobj=1` and aerial detections have `ddfobj=2` in the observation data.

### Mark-recapture distance sampling models

When using `mrds` models that include mark-recapture components (currently independent observer and trial modes are supported) then the format of the observation data needs to be checked to ensure that observations are not duplicated. The `observer` column is also required in the `observation.data`.

- *Independent observer mode* only unique observations (unique object IDs) are required.
- *Trial mode* only observations made by observer 1 are required.

---

dsm.cor

*Check for autocorrelation in residuals*

---

### Description

This function is deprecated, use `dsm_cor`.

### Usage

```
dsm.cor(
  dsm.obj,
  Transect.Label = "Transect.Label",
  Segment.Label = "Segment.Label",
  max.lag = 10,
  resid.type = "scaled.pearson",
  fun = cor,
  ylim = c(0, 1),
  subset = "all",
  ...
)
```

### Arguments

<code>dsm.obj</code>	a fitted <code>dsm</code> object.
<code>Transect.Label</code>	label for the transect (default: <code>Transect.Label</code> ). Using different labels can be useful when transects are split over geographical features or when transects are surveyed multiple times.
<code>Segment.Label</code>	label for the segments (default: <code>Segment.Label</code> ).The result of calling <code>order</code> must make sense.
<code>max.lag</code>	maximum lag to calculate at.

resid.type	the type of residuals used, see <a href="#">residuals.gam</a> . Defaults to "scaled.pearson" in the GAM case and "normalized" in the GAMM case (which are equivalent).
fun	the function to use, by default <a href="#">cor</a> , must take two column vectors as arguments.
ylim	user defined limits in y direction.
subset	which subset of the data should the correlation function be calculated on?
...	other options to pass to <a href="#">plot</a> .

---

 dsm.var.gam

*Prediction variance estimation assuming independence*


---

### Description

This function is deprecated, use [dsm\\_var\\_gam](#).

### Usage

```
dsm.var.gam(
  dsm.obj,
  pred.data,
  off.set,
  seglen.varname = "Effort",
  type.pred = "response"
)
```

### Arguments

dsm.obj	a model object fitted by <a href="#">dsm</a> .
pred.data	either: a single prediction grid or list of prediction grids. Each grid should be a <code>data.frame</code> with the same columns as the original data.
off.set	a a vector or list of vectors with as many elements as there are in <code>pred.data</code> . Each vector is as long as the number of rows in the corresponding element of <code>pred.data</code> . These give the area associated with each prediction cell. If a single number is supplied it will be replicated for the length of <code>pred.data</code> .
seglen.varname	name for the column which holds the segment length (default value "Effort").
type.pred	should the predictions be on the "response" or "link" scale? (default "response").

---

dsm.var.movblk	<i>Variance estimation via parametric moving block bootstrap</i>
----------------	--

---

## Description

This function is deprecated, use [dsm\\_var\\_movblk](#).

## Usage

```
dsm.var.movblk(
  dsm.object,
  pred.data,
  n.boot,
  block.size,
  off.set,
  ds.uncertainty = FALSE,
  samp.unit.name = "Transect.Label",
  progress.file = NULL,
  bs.file = NULL,
  bar = TRUE
)
```

## Arguments

dsm.object	object returned from <a href="#">dsm</a> .
pred.data	either: a single prediction grid or list of prediction grids. Each grid should be a data.frame with the same columns as the original data.
n.boot	number of bootstrap resamples.
block.size	number of segments in each block.
off.set	a a vector or list of vectors with as many elements as there are in pred.data. Each vector is as long as the number of rows in the corresponding element of pred.data. These give the area associated with each prediction cell. If a single number is supplied it will be replicated for the length of pred.data.
ds.uncertainty	incorporate uncertainty in the detection function? See Details, below. Note that this feature is EXPERIMENTAL at the moment.
samp.unit.name	name sampling unit to resample (default 'Transect.Label').
progress.file	path to a file to be used (usually by Distance) to generate a progress bar (default NULL – no file written).
bs.file	path to a file to store each bootstrap round. This stores all of the bootstrap results rather than just the summaries, enabling outliers to be detected and removed. (Default NULL).
bar	should a progress bar be printed to screen? (Default TRUE).

---

dsm.var.prop                      *Prediction variance propagation for DSMs*

---

### Description

This function is deprecated, use [dsm\\_var\\_prop](#).

### Usage

```
dsm.var.prop(
  dsm.obj,
  pred.data,
  off.set,
  seglen.varname = "Effort",
  type.pred = "response"
)
```

### Arguments

dsm.obj	a model object fitted by <a href="#">dsm</a> .
pred.data	either: a single prediction grid or list of prediction grids. Each grid should be a <code>data.frame</code> with the same columns as the original data.
off.set	a a vector or list of vectors with as many elements as there are in <code>pred.data</code> . Each vector is as long as the number of rows in the corresponding element of <code>pred.data</code> . These give the area associated with each prediction cell. If a single number is supplied it will be replicated for the length of <code>pred.data</code> .
seglen.varname	name for the column which holds the segment length (default value "Effort").
type.pred	should the predictions be on the "response" or "link" scale? (default "response").

---

dsm\_cor                              *Check for autocorrelation in residuals*

---

### Description

Once a DSM has been fitted to data, this function can be used to check for autocorrelation in the residuals.

### Usage

```
dsm_cor(
  dsm.obj,
  Transect.Label = "Transect.Label",
  Segment.Label = "Segment.Label",
  max.lag = 10,
```

```

    resid.type = "scaled.pearson",
    fun = cor,
    ylim = c(0, 1),
    subset = "all",
    ...
)

```

## Arguments

dsm.obj	a fitted dsm object.
Transect.Label	label for the transect (default: Transect.Label). Using different labels can be useful when transects are split over geographical features or when transects are surveyed multiple times.
Segment.Label	label for the segments (default: Segment.Label).The result of calling <a href="#">order</a> must make sense.
max.lag	maximum lag to calculate at.
resid.type	the type of residuals used, see <a href="#">residuals.gam</a> . Defaults to "scaled.pearson" in the GAM case and "normalized" in the GAMM case (which are equivalent).
fun	the function to use, by default <a href="#">cor</a> , must take two column vectors as arguments.
ylim	user defined limits in y direction.
subset	which subset of the data should the correlation function be calculated on?
...	other options to pass to <a href="#">plot</a> .

## Value

a plot or a vector of fun applied at the lags.

## Details

Within each Transect.Label, segments will be sorted according to their Segment.Labels. This may require some time to get right for your particular data. If one has multiple surveys where transects are revisited, for example, one may want to make Transect.Label a unique transect-survey identifier. Neither label need to be included in the model, they must just be present in the \$data field in the model. This usually means that they have to be in the segment data passed to [dsm](#).

The current iteration of this function will only plot correlations nicely, other things are up to you but you can get the function to return the data (by assigning the result to an object).

If there are NA values in the residuals then the correlogram will not be calculated. This usually occurs due to NA values in the covariates (so the smoother will not have fitted values there). Code like `any(is.na(dsm.obj$data))` might be helpful.

## Author(s)

David L. Miller

**Examples**

```

library(Distance)
library(dsm)

# load the data, see ?mexdolphins
data(mexdolphins)

# fit a model
hr.model <- ds(distdata, truncation=6000,
               key = "hr", adjustment = NULL)
mod1 <- dsm(count~s(x,y), hr.model, segdata, obsdata)

# look at lag 1 differences up to a maximum of lag 9, using deviance
# residuals
dsm_cor(mod1, resid.type="deviance", max.lag=9,
        Segment.Label="Sample.Label")

```

---

 dsm\_varprop

*Variance propagation for density surface models*


---

**Description**

Calculate the uncertainty in predictions from a fitted DSM, including uncertainty from the detection function.

**Usage**

```

dsm_varprop(
  model,
  newdata = NULL,
  trace = FALSE,
  var.type = "Vp",
  var_type = NULL
)

```

**Arguments**

model	a fitted <a href="#">dsm</a> .
newdata	the prediction grid. Set to NULL to avoid making predictions and just return model objects.
trace	for debugging, see how the scale parameter estimation is going.
var.type	which variance-covariance matrix should be used ("Vp" for variance-covariance conditional on smoothing parameter(s), "Vc" for unconditional). See <a href="#">gamObject</a> for an details/explanation. If in doubt, stick with the default, "Vp".
var_type	deprecated, use var.type instead.

## Details

When we make predictions from a spatial model, we also want to know the uncertainty about that abundance estimate. Since density surface models are 2 (or more) stage models, we need to incorporate the uncertainty from the earlier stages (i.e. the detection function) into our "final" uncertainty estimate.

This function will refit the spatial model but include the Hessian of the offset as an extra term. Variance estimates using this new model can then be used to calculate the variance of predicted abundance estimates which incorporate detection function uncertainty. Importantly this requires that if the detection function has covariates, then these do not vary within a segment (so, for example covariates like sex cannot be used).

For more information on how to construct the prediction grid data.frame, newdata, see [predict.dsm](#).

This routine is only useful if a detection function with covariates has been used in the DSM.

Note that we can use `var.type="Vc"` here (see `gamObject`), which is the variance-covariance matrix for the spatial model, corrected for smoothing parameter uncertainty. See Wood, Pya & Sørfer (2016) for more information.

Models with fixed scale parameters (e.g., negative binomial) do not require an extra round of optimisation.

## Value

a list with elements:

- `old_model` fitted model supplied to the function as `model`
- `refit` refitted model object, with extra term
- `pred` point estimates of predictions at `newdata`
- `var` total variance calculated over all of `newdata`
- `ses` standard error for each prediction cell in `newdata` if `newdata=NULL` then the last three entries are NA.

## Diagnostics

The summary output from the function includes a simple diagnostic that shows the average probability of detection from the "original" fitted model (the model supplied to this function; column `Fitted.model`) and the probability of detection from the refitted model (used for variance propagation; column `Refitted.model`) along with the standard error of the probability of detection from the fitted model (`Fitted.model.se`), at the unique values of any factor covariates used in the detection function (for continuous covariates the 5%, 50% and 95% quantiles are shown). If there are large differences between the probabilities of detection then there are potentially problems with the fitted model, the variance propagation or both. This can be because the fitted model does not account for enough of the variability in the data and in refitting the variance model accounts for this in the random effect.

## Author(s)

David L. Miller, based on code from Mark V. Bravington and Sharon L. Hedley.

## References

- Bravington, M. V., Miller, D. L., & Hedley, S. L. (2021). Variance Propagation for Density Surface Models. *Journal of Agricultural, Biological and Environmental Statistics*. <https://doi.org/10.1007/s13253-021-00438-2>
- Williams, R., Hedley, S.L., Branch, T.A., Bravington, M.V., Zerbini, A.N. and Findlay, K.P. (2011). Chilean Blue Whales as a Case Study to Illustrate Methods to Estimate Abundance and Evaluate Conservation Status of Rare Species. *Conservation Biology* 25(3), 526-535.
- Wood, S.N., Pya, N. and Säfken, B. (2016) Smoothing parameter and model selection for general smooth models. *Journal of the American Statistical Association*, 1-45.

---

dsm\_var\_gam

*Prediction variance estimation assuming independence*

---

## Description

If one is willing to assume the the detection function and spatial model are independent, this function will produce estimates of variance of predictions of abundance, using the result that squared coefficients of variation will add.

## Usage

```
dsm_var_gam(
  dsm.obj,
  pred.data,
  off.set,
  seglen.varname = "Effort",
  type.pred = "response"
)
```

## Arguments

- |                |  |
|----------------|--|
| dsm.obj        | a model object fitted by <a href="#">dsm</a> .   |
| pred.data      | either: a single prediction grid or list of prediction grids. Each grid should be a <code>data.frame</code> with the same columns as the original data.  |
| off.set        | a a vector or list of vectors with as many elements as there are in <code>pred.data</code> . Each vector is as long as the number of rows in the corresponding element of <code>pred.data</code> . These give the area associated with each prediction cell. If a single number is supplied it will be replicated for the length of <code>pred.data</code> . |
| seglen.varname | name for the column which holds the segment length (default value "Effort").   |
| type.pred      | should the predictions be on the "response" or "link" scale? (default "response").   |



**Value**

a list with elements

- model the fitted model object
- pred.var variance of the regions given in pred.data.
- bootstrap logical, always FALSE
- model the fitted model with the extra term
- dsm.object the original model (dsm.obj above)

**Author(s)**

David L. Miller

**Examples**

```
## Not run:
library(Distance)
library(dsm)

# load the Gulf of Mexico dolphin data (see ?mexdolphins)
data(mexdolphins)

# fit a detection function and look at the summary
hr.model <- ds(distdata, truncation=6000,
              key = "hr", adjustment = NULL)
summary(hr.model)

# fit a simple smooth of x and y
mod1 <- dsm(count~s(x, y), hr.model, segdata, obsdata)

# Calculate the variance
# this will give a summary over the whole area in mexdolphins$preddata
mod1.var <- dsm_var_gam(mod1, preddata, off.set=preddata$area)

## End(Not run)
```

---

dsm\_var\_movblk

*Variance estimation via parametric moving block bootstrap*

---

**Description**

Estimate the variance in abundance over an area using a moving block bootstrap. Two procedures are implemented, one incorporating detection function uncertainty, one not.

**Usage**

```

dsm_var_movblk(
  dsm.object,
  pred.data,
  n.boot,
  block.size,
  off.set,
  ds.uncertainty = FALSE,
  samp.unit.name = "Transect.Label",
  progress.file = NULL,
  bs.file = NULL,
  bar = TRUE
)

```

**Arguments**

dsm.object	object returned from <a href="#">dsm</a> .
pred.data	either: a single prediction grid or list of prediction grids. Each grid should be a <code>data.frame</code> with the same columns as the original data.
n.boot	number of bootstrap resamples.
block.size	number of segments in each block.
off.set	a a vector or list of vectors with as many elements as there are in <code>pred.data</code> . Each vector is as long as the number of rows in the corresponding element of <code>pred.data</code> . These give the area associated with each prediction cell. If a single number is supplied it will be replicated for the length of <code>pred.data</code> .
ds.uncertainty	incorporate uncertainty in the detection function? See Details, below. Note that this feature is EXPERIMENTAL at the moment.
samp.unit.name	name sampling unit to resample (default 'Transect.Label').
progress.file	path to a file to be used (usually by Distance) to generate a progress bar (default NULL – no file written).
bs.file	path to a file to store each bootstrap round. This stores all of the bootstrap results rather than just the summaries, enabling outliers to be detected and removed. (Default NULL).
bar	should a progress bar be printed to screen? (Default TRUE).

**Details**

Setting `ds.uncertainty=TRUE` will incorporate detection function uncertainty directly into the bootstrap. This is done by generating observations from the fitted detection function and then re-fitting a new detection function (of the same form), then calculating a new effective strip width. Rejection sampling is used to generate the observations (except in the half-normal case) so the procedure can be rather slow. Note that this is currently not supported with covariates in the detection function.

Setting `ds.uncertainty=FALSE` will incorporate detection function uncertainty using the delta method. This assumes that the detection function and the spatial model are INDEPENDENT. This is probably not reasonable.

**Examples**

```
## Not run:
library(Distance)
library(dsm)

# load the Gulf of Mexico dolphin data (see ?mexdolphins)
data(mexdolphins)

# fit a detection function and look at the summary
hr.model <- ds(distdata, truncation=6000,
              key = "hr", adjustment = NULL)
summary(hr.model)

# fit a simple smooth of x and y
mod1 <- dsm(count~s(x, y), hr.model, segdata, obsdata)
summary(mod1)

# calculate the variance by 500 moving block bootstraps
mod1.movblk <- dsm_var_movblk(mod1, preddata, n.boot = 500,
                             block.size = 3, samp.unit.name = "Transect.Label",
                             off.set = preddata$area,
                             bar = TRUE, bs.file = "mexico-bs.csv", ds.uncertainty = TRUE)

## End(Not run)
```

---

dsm\_var\_prop

*Prediction variance propagation for DSMs*


---

**Description**

To ensure that uncertainty from the detection function is correctly propagated to the final variance estimate of abundance, this function uses a method first detailed in Williams et al (2011), further explanation is given in Bravington et al. (2021).

**Usage**

```
dsm_var_prop(
  dsm.obj,
  pred.data,
  off.set,
  seglen.varname = "Effort",
  type.pred = "response"
)
```

**Arguments**

dsm.obj	a model object fitted by <a href="#">dsm</a> .
pred.data	either: a single prediction grid or list of prediction grids. Each grid should be a data.frame with the same columns as the original data.

<code>off.set</code>	a a vector or list of vectors with as many elements as there are in <code>pred.data</code> . Each vector is as long as the number of rows in the corresponding element of <code>pred.data</code> . These give the area associated with each prediction cell. If a single number is supplied it will be replicated for the length of <code>pred.data</code> .
<code>seglen.varname</code>	name for the column which holds the segment length (default value "Effort").
<code>type.pred</code>	should the predictions be on the "response" or "link" scale? (default "response").

### Details

The idea is to refit the spatial model but including an extra random effect. This random effect has zero mean and hence to effect on point estimates. Its variance is the Hessian of the detection function. Variance estimates then incorporate detection function uncertainty. Further mathematical details are given in the paper in the references below.

Many prediction grids can be supplied by supplying a list of `data.frames` to the function.

Note that this routine simply calls `dsm_varprop`. If you don't require multiple prediction grids, the other routine will probably be faster.

This routine is only useful if a detection function with covariates has been used in the DSM.

### Value

a list with elements

- `model` the fitted model object
- `pred.var` variance of each region given in `pred.data`
- `bootstrap` logical, always FALSE
- `pred.data` as above
- `off.set` as above
- `model` the fitted model with the extra term
- `dsm.object` the original model, as above
- `model.check` simple check of subtracting the coefficients of the two models to see if there is a large difference
- `deriv` numerically calculated Hessian of the offset

### Diagnostics

The summary output from the function includes a simply diagnostic that shows the average probability of detection from the "original" fitted model (the model supplied to this function; column `Fitted.model`) and the probability of detection from the refitted model (used for variance propagation; column `Refitted.model`) along with the standard error of the probability of detection from the fitted model (`Fitted.model.se`), at the unique values of any factor covariates used in the detection function (for continuous covariates the 5%, 50% and 95% quantiles are shown). If there are large differences between the probabilities of detection then there are potentially problems with the fitted model, the variance propagation or both. This can be because the fitted model does not account for enough of the variability in the data and in refitting the variance model accounts for this in the random effect.

**Limitations**

Note that this routine is only useful if a detection function has been used in the DSM. It cannot be used when the `abundance.est` or `density.est` responses are used. Importantly this requires that if the detection function has covariates, then these do not vary within a segment (so, for example covariates like sex cannot be used).

**Author(s)**

Mark V. Bravington, Sharon L. Hedley. Bugs added by David L. Miller.

**References**

- Bravington, M. V., Miller, D. L., & Hedley, S. L. (2021). Variance Propagation for Density Surface Models. *Journal of Agricultural, Biological and Environmental Statistics*. <https://doi.org/10.1007/s13253-021-00438-2>
- Williams, R., Hedley, S.L., Branch, T.A., Bravington, M.V., Zerbini, A.N. and Findlay, K.P. (2011). Chilean Blue Whales as a Case Study to Illustrate Methods to Estimate Abundance and Evaluate Conservation Status of Rare Species. *Conservation Biology* 25(3), 526-535.

---

 dummy\_ddf

---

*Detection function objects when detection is certain*


---

**Description**

Create a detection function object for strip/plot surveys for use in density surface models.

**Usage**

```
dummy_ddf(object, size = 1, width, left = 0, transect = "line")
```

**Arguments**

object	numeric vector of object identifiers, relating to the object field in the observation data of the DSM.
size	group size for each observation (default all groups size 1)
width	right truncation
left	left truncation (default 0, no left truncation)
transect	"line" or "point" transect

**Author(s)**

David L Miller

generate.ds.uncertainty

*Generate data from a fitted detection function*

---

### Description

When using `dsm.var.movblk` if `ds.uncertainty=TRUE`, this procedure generates data from the fitted detection function (assuming that it is correct).

### Usage

```
generate.ds.uncertainty(ds.object)
```

### Arguments

`ds.object` a fitted detection function object (as returned by a call to `ddf.ds`).

### Note

This function changes the random number generator seed. To avoid any potential side-effects, use something like: `seed <- get(".Random.seed", envir=.GlobalEnv)` before running code and `assign(".Random.seed", seed, envir=.GlobalEnv)` after.

### Author(s)

David L. Miller

---

generate.mb.sample

*Moving block bootstrap sampler*

---

### Description

Not usually used on its own, called from within `dsm.var.movblk`.

### Usage

```
generate.mb.sample(  
  num.blocks.required,  
  block.size,  
  which.blocks,  
  dsm.data,  
  unit.info,  
  n.units  
)
```

**Arguments**

num.blocks.required	number of blocks that we need.
block.size	number of segments per block.
which.blocks	which blocks should be sampled.
dsm.data	the \$data element of the result of a call to <code>dsm</code> .
unit.info	result of calling <code>block.info.per.su</code> .
n.units	number of sampling units.

**Value**

vector of log-residuals

---

latlong2km	<i>Convert latitude and longitude to Northings and Eastings</i>
------------	---

---

**Description**

Convert longitude and latitude co-ordinates to kilometres west-east and south-north from axes through (lon0,lat0) using the "spherical law of cosines".

**Usage**

```
latlong2km(lon, lat, lon0 = sum(range(lon))/2, lat0 = sum(range(lat))/2)
```

**Arguments**

lon	longitude
lat	latitude
lon0	longitude reference point (defaults to mean longitude)
lat0	latitude reference point (defaults to mean latitude)

**Details**

**WARNING:** This is an approximate procedure for converting between latitude/ longitude and Northing/Easting. Consider using projection conversions available in packages `sp`, `sf` and `rgdal` for better results.

**Value**

list with elements `km.e` and `km.n`.

**Author(s)**

Simon N. Wood

`make.soapgrid`*Create a knot grid for the internal part of a soap film smoother.*

---

**Description**

This routine simply creates a grid of knots (in the correct format) to be used as in the "internal" part of the soap film smoother

**Usage**

```
make.soapgrid(bnd, n.grid)
```

**Arguments**

<code>bnd</code>	list with elements <code>x</code> and <code>y</code> which give the locations of the boundary vertices. The first and last elements should be the same.
<code>n.grid</code>	either one number giving the number of points along the <code>x</code> and <code>y</code> axes that should be used to create the grid, or a vector giving the number in the <code>x</code> direction, then <code>y</code> direction.

**Value**

a list with elements `x` and `y`, containing the knot locations.

**Author(s)**

David L Miller

---

`mexdolphins`*Pan-tropical spotted dolphins in the Gulf of Mexico*

---

**Description**

Data from a combination of several NOAA shipboard surveys conducted on pan-tropical spotted dolphins in the Gulf of Mexico. The data consist of 47 observations of groups of dolphins. The group size was recorded, as well as the Beaufort sea state at the time of the observation. Coordinates for each observation and bathymetry data were also available as covariates for the analysis. A complete example analysis (and description of the data) is provided at <http://distancesampling.org/R/vignettes/mexico-analysis.html>.



## References

Halpin, P.N., A.J. Read, E. Fujioka, B.D. Best, B. Donnelly, L.J. Hazen, C. Kot, K. Urian, E. LaBrecque, A. Dimatteo, J. Cleary, C. Good, L.B. Crowder, and K.D. Hyrenbach. 2009. OBIS-SEAMAP: The world data center for marine mammal, sea bird, and sea turtle distributions. *Oceanography* 22(2):104-115

NOAA Southeast Fisheries Science Center. 1996. Report of a Cetacean Survey of Oceanic and Selected Continental Shelf Waters of the Northern Gulf of Mexico aboard NOAA Ship Oregon II (Cruise 220)

---

 obs\_exp

---

*Observed versus expected diagnostics for fitted DSMs*


---

## Description

Given a covariate, calculate the observed and expected counts for each unique value of the covariate. This can be a useful goodness of fit check for DSMs.

## Usage

```
obs_exp(model, covar, cut = NULL)
```

## Arguments

model	a fitted <a href="#">dsm</a> model object
covar	covariate to aggregate by (character)
cut	vector of cut points to aggregate at. If not supplied, the unique values of covar are used.

## Details

One strategy for model checking is to calculate observed and expected counts at different aggregations of the variable. If these match well then the model fit is good.

## Value

data.frame with values of observed and expected counts.

## Author(s)

David L Miller, on the suggestion of Mark Bravington.

**Examples**

```
## Not run:
library(Distance)
library(dsm)

# example with the Gulf of Mexico dolphin data
data(mexdolphins)
hr.model <- ds(distdata, truncation=6000,
              key = "hr", adjustment = NULL)
mod1 <- dsm(count~s(x,y), hr.model, segdata, obsdata)

## End(Not run)
```

---

plot.dsm

*Plot a density surface model.*

---

**Description**

See [plot.gam](#).

**Usage**

```
## S3 method for class 'dsm'
plot(x, ...)
```

**Arguments**

x                    a model fitted by [dsm](#)  
...                   other arguments passed to [plot.gam](#).

**Value**

a plot!

**Author(s)**

David L. Miller

**See Also**

[dsm](#) [plot.gam](#)

---

plot.dsm.var                      *Create plots of abundance uncertainty*

---

### Description

Note that the prediction data set must have x and y columns even if these were not used in the model.

### Usage

```
## S3 method for class 'dsm.var'
plot(
  x,
  poly = NULL,
  limits = NULL,
  breaks = NULL,
  legend.breaks = NULL,
  xlab = "x",
  ylab = "y",
  observations = TRUE,
  plot = TRUE,
  boxplot.coef = 1.5,
  x.name = "x",
  y.name = "y",
  gg.grad = NULL,
  ...
)
```

### Arguments

x	a dsm.var object
poly	a list or data.frame with columns x and y, which gives the coordinates of a polygon to draw. It may also optionally have a column group, if there are many polygons.
limits	limits for the fill colours
breaks	breaks for the colour fill
legend.breaks	breaks as they should be displayed
xlab	label for the x axis
ylab	label for the y axis
observations	should observations be plotted?
plot	actually plot the map, or just return a ggplot2 object?
boxplot.coef	control trimming (as in <a href="#">summary.dsm.var</a> ), only has an effect if the bootstrap file was saved.
x.name	name of the variable to plot as the x axis.

y.name	name of the variable to plot as the y axis.
gg.grad	optional <a href="#">ggplot</a> gradient object.
...	any other arguments

**Value**

a plot

**Details**

In order to get plotting to work with [dsm\\_var\\_prop](#) and [dsm\\_var\\_gam](#), one must first format the data correctly since these functions are designed to compute very general summaries. One summary is calculated for each element of the list `pred` supplied to [dsm\\_var\\_prop](#) and [dsm\\_var\\_gam](#).

For a plot of uncertainty over a prediction grid, `pred` (a `data.frame`), say, we can create the correct format by simply using `pred.new <- split(pred, 1:nrow(pred))`.

**Author(s)**

David L. Miller

**See Also**

[dsm\\_var\\_prop](#), [dsm\\_var\\_gam](#), [dsm\\_var\\_movblk](#)

---

plot_pred_by_term	<i>Spatially plot predictions per model term</i>
-------------------	--

---

**Description**

Plot the effect of each smooth in the model spatially. For each term in the model, plot its effect in space. Plots are made on the same scale, so that the relative influence of each smooth can be seen.

**Usage**

```
plot_pred_by_term(dsm.obj, data, location.cov = c("x", "y"))
```

**Arguments**

dsm.obj	fitted <a href="#">dsm</a> object
data	data to use to plot (often the same as the prediction grid), data should also include width and height columns for plotting
location.cov	deprecated, use <code>location.cov</code>

**Value**

a `ggplot2` plot

**Author(s)**

David L Miller (idea taken from inlabru)

**Examples**

```
## Not run:
library(Distance)
library(dsm)

# load the Gulf of Mexico dolphin data and fit a model
data(mexdolphins)
hr.model <- ds(distdata, truncation=6000,
              key = "hr", adjustment = NULL)
mod1 <- dsm(count~s(x,y) + s(depth), hr.model, segdata, obsdata)

preddata$width <- preddata$height <- sqrt(preddata$area)

# make the plot
plot_pred_by_term(mod1, preddata, c("x", "y"))

# better plot would be
# library(viridis)
# plot_pred_by_term(mod1, preddata, c("x", "y")) + scale_fill_viridis()

## End(Not run)
```

---

predict.dsm

*Predict from a fitted density surface model*

---

**Description**

Make predictions of density or abundance outside (or inside) the covered area.

**Usage**

```
## S3 method for class 'dsm'
predict(object, newdata = NULL, off.set = NULL, type = "response", ...)
```

**Arguments**

object	a fitted <code>dsm</code> object
newdata	spatially referenced covariates e.g. altitude, depth, distance to shore, etc. Covariates in the <code>data.frame</code> must have names <i>identical</i> to variable names used in fitting the model
off.set	area of each of the cells in the prediction grid. Should be in the same units as the segments/distances given to <code>dsm</code> . Replaces the column in <code>newdata</code> called <code>off.set</code> if it is supplied. Ignored if <code>newdata</code> is not supplied

type            what scale should the results be on. The default is "response", see [predict.gam](#) for an explanation of other options (usually not necessary)

...            any other arguments passed to [predict.gam](#)

### Details

If newdata is not supplied, predictions are made for the data that built the model. Note that the order of the results will not necessarily be the same as the segdata (segment data) data.frame that was supplied to [dsm](#).

The area.off.set is used if that argument is supplied, otherwise it will look for the areas in the column named off.set in the newdata. Either way the link function (usually log) will be applied to the offsets, so there is no need to log them before passing them to this function.

### Value

predicted values on the response scale by default (unless type is specified, in which case see [predict.gam](#)).

### Author(s)

David L. Miller

### See Also

[predict.gam](#), [dsm\\_var\\_gam](#), [dsm\\_var\\_prop](#)

---

predict.fake\_ddf            *Prediction for fake detection functions*

---

### Description

Prediction function for dummy detection functions. The function returns as many 1s as there are rows in newdata. If esw=TRUE then the strip width is returned.

### Usage

```
## S3 method for class 'fake_ddf'
predict(
  object,
  newdata = NULL,
  compute = FALSE,
  int.range = NULL,
  esw = FALSE,
  ...
)
```

**Arguments**

object	model object
newdata	how many 1s should we return?
compute	unused, compatibility with <a href="#">mrds::predict</a>
int.range	unused, compatibility with <a href="#">mrds::predict</a>
esw	should the strip width be returned?
...	for S3 consistency

**Author(s)**

David L Miller

---

print.dsm	<i>Print a description of a density surface model object</i>
-----------	--

---

**Description**

This method just gives a short description of the fitted model. Use the [summary.dsm](#) method for more information.

**Usage**

```
## S3 method for class 'dsm'
print(x, ...)
```

**Arguments**

x	a model fitted by <a href="#">dsm</a>
...	unspecified and unused arguments for S3 consistency

**Value**

NULL

**Author(s)**

David L. Miller

---

<code>print.dsm.var</code>	<i>Print a description of a density surface model variance object</i>
----------------------------	---

---

**Description**

This method only provides a short summary, use the [summary.dsm.var](#) method for information.

**Usage**

```
## S3 method for class 'dsm.var'  
print(x, ...)
```

**Arguments**

<code>x</code>	a dsm variance object
<code>...</code>	unspecified and unused arguments for S3 consistency

**Value**

NULL

**Author(s)**

David L. Miller

**See Also**

[summary.dsm.var](#)

---

<code>print.dsm_varprop</code>	<i>Print a description of a density surface model variance object</i>
--------------------------------	---

---

**Description**

This method only provides a short summary, see [summary.dsm\\_varprop](#).

**Usage**

```
## S3 method for class 'dsm_varprop'  
print(x, ...)
```

**Arguments**

<code>x</code>	a dsm variance object
<code>...</code>	unspecified and unused arguments for S3 consistency



**Author(s)**

David L. Miller

**See Also**

[summary.dsm\\_varprop](#)

---

`print.summary.dsm.var` *Print summary of density surface model variance object*

---

**Description**

See [summary.dsm.var](#) for information.

**Usage**

```
## S3 method for class 'summary.dsm.var'  
print(x, ...)
```

**Arguments**

<code>x</code>	a summary of dsm variance object
<code>...</code>	unspecified and unused arguments for S3 consistency

**Value**

NULL

**Author(s)**

David L. Miller

**See Also**

[summary.dsm.var](#)

---

```
print.summary.dsm_varprop
      Print summary of density surface model variance object
```

---

**Description**

See [summary.dsm\\_varprop](#) for information.

**Usage**

```
## S3 method for class 'summary.dsm_varprop'
print(x, ...)
```

**Arguments**

x	a summary of dsm variance object
...	unspecified and unused arguments for S3 consistency

**Value**

NULL

**Author(s)**

David L. Miller

**See Also**

[summary.dsm.var](#)

---

```
rqgam.check      Randomised quantile residuals check plot for GAMs/DSMs
```

---

**Description**

This function is deprecated, use [rqgam\\_check](#).

**Usage**

```
rqgam.check(gam.obj, ...)
```

**Arguments**

gam.obj	a <a href="#">gam</a> , <a href="#">glm</a> or <a href="#">dsm</a> object.
...	arguments passed on to all plotting functions

---

`rqgam_check`*Randomised quantile residuals check plot for GAMs/DSMs*

---

### Description

Reproduces the "Resids vs. linear pred" plot from `gam.check` but using randomised quantile residuals, a la Dunn and Smyth (1996). Checks for heteroskedasticity as usual, looking for "funnel"-type structures in the points, which is much easier with randomised quantile residuals than with deviance residuals, when your model uses a count distribution as the response.

### Usage

```
rqgam_check(gam.obj, ...)
```

### Arguments

<code>gam.obj</code>	a <code>gam</code> , <code>glm</code> or <code>dsm</code> object.
<code>...</code>	arguments passed on to all plotting functions

### Details

Note that this function only works with negative binomial and Tweedie response distributions.

Earlier versions of this function produced the full `gam.check` output, but this was confusing as only one of the plots was really useful. Checks of `k` are not computed, these need to be done using `gam.check`.

### Value

just plots!

### Author(s)

Based on code by Natalie Kelly, bugs added by Dave Miller

### Examples

```
library(Distance)
library(dsm)
library(tweedie)

# load the Gulf of Mexico dolphin data (see ?mexdolphins)
data(mexdolphins)

# fit a detection function and look at the summary
hr.model <- ds(distdata, truncation=6000,
              key = "hr", adjustment = NULL)

# fit a simple smooth of x and y with a Tweedie response with estimated
```

```
# p parameter
mod1 <- dsm(count~s(x, y), hr.model, segdata, obsdata, family=tw())
rqgam_check(mod1)
```

---

summary.dsm

*Summarize a fitted density surface model*


---

### Description

Gives a brief summary of a fitted [dsm](#) object.

### Usage

```
## S3 method for class 'dsm'
summary(object, ...)
```

### Arguments

object            a model fitted by [dsm](#)  
 ...                other arguments passed to [summary.gam](#)

### Value

a summary object

### Author(s)

David L. Miller

### See Also

[dsm](#)

---

summary.dsm.var

*Summarize the variance of a density surface model*


---

### Description

Gives a brief summary of a fitted dsm variance object.

**Usage**

```
## S3 method for class 'dsm.var'
summary(
  object,
  alpha = 0.05,
  boxplot.coef = 1.5,
  bootstrap.subregions = NULL,
  ...
)
```

**Arguments**

object	a dsm.var object
alpha	alpha level for confidence intervals (default 0.05 to give a 95\ confidence intervals)
boxplot.coef	the value of coef used to calculate the outliers see <a href="#">boxplot</a> .
bootstrap.subregions	list of vectors of logicals or indices for subregions for which variances need to be calculated (only for bootstraps (see <a href="#">dsm.var.prop</a> for how to use subregions with variance propagation).
...	unused arguments for S3 compatibility

**Value**

a summary object

**Author(s)**

David L. Miller

**See Also**

[dsm.var.movblk](#), [dsm.var.prop](#)

---

summary.dsm\_varprop     *Summarize the variance of a density surface model*

---

**Description**

Gives a brief summary of a fitted [dsm\\_varprop](#) variance object.

**Usage**

```
## S3 method for class 'dsm_varprop'
summary(object, alpha = 0.05, ...)
```

**Arguments**

object            a dsm.var object  
 alpha            alpha level for confidence intervals (default 0.05 to give a 95% confidence interval)  
 ...               unused arguments for S3 compatibility

**Value**

a summary object

**Author(s)**

David L. Miller

**See Also**

[dsm\\_varprop](#), [summary.dsm.var](#)

---

trim.var

*Trimmed variance*

---

**Description**

Trim the variance estimates from the bootstrap. This is defined as the percentage defined as amount necessary to bring median and trimmed mean within 8% of each other these are defined as 'outliers'.

**Usage**

```
trim.var(untrimmed.bootstraps, boxplot.coef = 1.5)
```

**Arguments**

untrimmed.bootstraps  
    (usually the \$study.area.total element of a returned [dsm.var.movblk](#) bootstrap object.  
 boxplot.coef        the value of coef used to calculate the outliers see [boxplot](#).

**Value**

trimmed variance

**Author(s)**

Louise Burt

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vis.concurvity	<i>Visualise concurvity between terms in a GAM</i>
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**Description**

This function is deprecated, use [vis\\_concurvity](#).

**Usage**

```
vis.concurvity(model, type = "estimate")
```

**Arguments**

model	fitted model
type	concurvity measure to plot, see <a href="#">concurvity</a>

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vis_concurvity	<i>Visualise concurvity between terms in a GAM</i>
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**Description**

Plot measures of how much one term in the model could be explained by another. When values are high, one should consider re-running variable selection with one of the offending variables removed to check for stability in term selection.

**Usage**

```
vis_concurvity(model, type = "estimate")
```

**Arguments**

model	fitted model
type	concurvity measure to plot, see <a href="#">concurvity</a>

**Details**

These methods are considered somewhat experimental at this time. Consult [concurvity](#) for more information on how concurvity measures are calculated.

**Author(s)**

David L Miller

**Examples**

```
## Not run:
library(Distance)
library(dsm)

# load the Gulf of Mexico dolphin data (see ?mexdolphins)
data(mexdolphins)

# fit a detection function and look at the summary
hr.model <- ds(distdata, truncation=6000,
              key = "hr", adjustment = NULL)

# fit a simple smooth of x and y to counts
mod1 <- dsm(count~s(x,y)+s(depth), hr.model, segdata, obsdata)

# visualise concurvity using the "estimate" metric
vis_concurvity(mod1)

## End(Not run)
```



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